

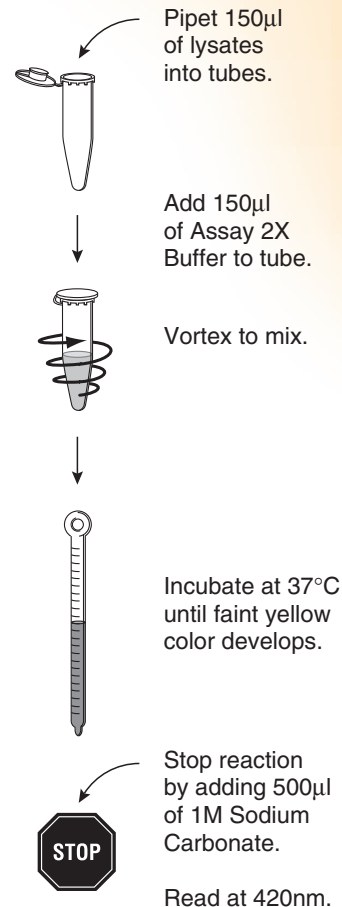
# $\beta$ -Galactosidase Enzyme Assay System with Reporter Lysis Buffer

INSTRUCTIONS FOR USE OF PRODUCT E2000.

*Quick*  
PROTOCOL

## Standard Protocol (96-well plate protocol on reverse side)

1. Prepare cell lysates (see Section 3 of the Technical Bulletin).
2. Thaw system components and mix well. Place 2X Assay Buffer on ice.
3. It may be necessary to dilute cell lysates in 1X Reporter Lysis Buffer (RLB). A 2:1 dilution of lysate to 1X RLB (100 $\mu$ l of lysate plus 50 $\mu$ l of 1X RLB) is a good starting dilution, but up to 150 $\mu$ l of cell lysate can be used per reaction.
4. Prepare a negative control (lysate from nontransfected cells) in 1X RLB, using the same dilution as in Step 3.
5. Add 150 $\mu$ l of Assay 2X Buffer to each tube. Mix by vortexing briefly.
6. Incubate reactions at 37°C for 30 minutes or until a faint yellow color has developed.
7. Stop reactions by adding 500 $\mu$ l of 1M Sodium Carbonate. Vortex briefly to mix.
8. Read the absorbance at 420nm.



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For additional protocol information, see Technical Bulletin #TB097, available online at: [www.promega.com/tbs](http://www.promega.com/tbs)

### ORDERING/TECHNICAL INFORMATION:

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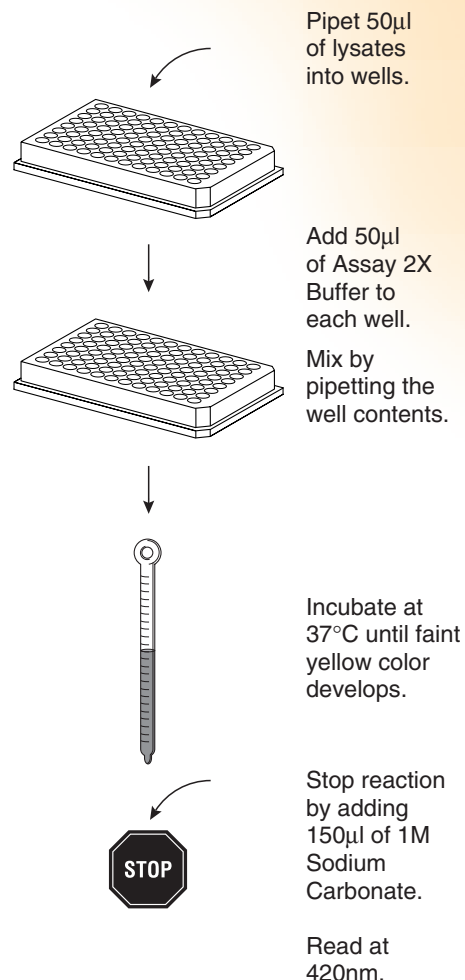
# $\beta$ -Galactosidase Enzyme Assay System with Reporter Lysis Buffer

INSTRUCTIONS FOR USE OF PRODUCT E2000.

**Quick**  
PROTOCOL

## 96-Well Plate Assay

1. Prepare cell lysates (see Section 3 of the Technical Bulletin).
2. Thaw system components and mix well. Place 2X Assay Buffer on ice.
3. It may be necessary to dilute cell lysates in 1X Reporter Lysis Buffer (RLB). Mix 30 $\mu$ l of lysate with 20 $\mu$ l of 1X RLB, as a starting dilution (as much as 50 $\mu$ l of cell lysate can be used per reaction). Pipet 50 $\mu$ l of cell lysates into wells of a 96-well plate.
4. Prepare a negative control, using nontransfected cells, in 1X RLB at the same dilution as used in Step 3.
5. Add 50 $\mu$ l of Assay 2X Buffer to each well.
6. Mix samples by pipetting the contents of each well. Cover the plate.
7. Incubate the plate at 37°C for 30 minutes or until a faint yellow color has developed. Due to the small sample volumes in the plate, do not incubate reactions overnight.
8. Stop reactions by adding 150 $\mu$ l of 1M Sodium Carbonate. Mix by pipetting the contents of each well gently to avoid bubbles.
9. Read the absorbance at 420nm.



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